



BoneTRAP[®] (TRAcP 5b) ELISA

SB-TR201R

Research Use Only

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BoneTRAP[®] (TRAcP 5b) ELISA

Test for the quantitative determination of the active isoform 5b of the tartrate-resistant acid phosphatase (TRACP)

Cat. no.: SB-TR201R

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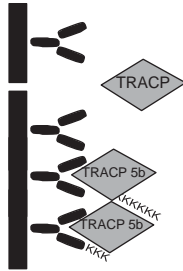
Introduction

High amount of tartrate-resistant acid phosphatase (TRACP) is expressed by bone-resorbing osteoclasts and activated macrophages (1). Two forms of TRACP circulate in human blood, known as TRACP 5a and TRACP 5b (2). TRACP 5b is derived from osteoclasts and TRACP 5a from macrophages (3).

Osteoclasts secrete TRACP 5b into the blood circulation as an active enzyme that is inactivated and degraded to fragments before it is removed from the circulation. Thus, TRACP 5b activity does not accumulate into the circulation in renal or hepatic failure (4,5). All serum TRACP 5b activity is derived from osteoclasts. Diurnal variability of serum TRACP 5b activity is low and the levels are not affected by feeding, allowing sample collection at any time of day (5).

[®]
The BoneTRAP[®] (TRAcP 5b) ELISA assay is to be used for research use only and is not to be used to aid in any clinical diagnosis.

Test principle



The plate is coated with anti-TRACP antibodies (mono-clonal).

Calibrators, Control and patient samples are added. Releasing reagent is added.

Dissociation of active TRACP 5b from the binding proteins.

TRACP 5b is bound by the anti-TRACP antibodies. Incubation with pNPP substrate (*).

The reaction is stopped by adding sodium hydroxide. The absorption is read photometrically.

Advantages of the test

- ✓ Measures TRACP 5b activity that is released specifically from osteoclasts.
- ✓ No interference with TRACP 5a or other phosphatases.
- ✓ Hemolysis has no effect on results.
- ✓ No diurnal variation.
- ✓ Not affected by functional disorders of kidney and liver.
- ✓ No dietary influences.

KIT CONTENTS

Cat. no.: SB-TR201A

1. **MICROPLAT** Microplate: 12 x 8 wells (with frame and desiccant in aluminium bag), F-form, coated with monoclonal anti-TRACP antibody (mouse) and BSA, ready to use.
2. **CTRL** Controls: 2 x 2 vials with 0.5 ml each, human recombinant TRACP, lyophilized, Xn, Harmful, R 22-52/53, S 36-60, contains < 1 % sodium azide and BSA.
3. **CAL** Calibrators: 2 x 6 vials with 0.5 ml each, human recombinant TRACP, lyophilized, Xn, Harmful, R 22-52/53, S 36-60, contains < 1 % sodium azide and BSA. The exact value of each calibrator is printed on the Quality Control Report.

4. **WASHBUF 25X** Wash Buffer: 1 bottle with 40 ml TBS/Tween (25x conc.), pH 7.65 – 7.85, contains < 1 % Germall®II.
5. **SAMPDIL** Sample Diluent: 1 vial with 15 ml, sodium chloride solution, ready to use, contains < 1 % Germall®II.
6. **RELEASESREAG** Releasing Reagent: 1 vial with 8 ml, pH 6.9 – 7.1, ready to use, contains < 1 % Germall®II.
7. **SUBSBUF** Substrate Buffer: 2 vials with 10 ml each, sodium acetate buffer, pH 5.95 – 6.05, ready to use, contains < 1 % Germall®II.
8. **SUBS pNPP** Substrate Tablets: 4 tablets, contain p-nitrophenyl phosphate (pNPP).
9. **NaOH** Stop Solution: 1 vial with 6 ml, 0.32 M sodium hydroxide, ready to use, Xi, Irritant, R 36/38, S 26-37-60.

1. STORAGE AND STABILITY

MATERIAL/REAGENT	STATE	STORAGE	STABILITY
Test kit	unopened	2...8 °C	until expiry date
Microplate	opened	2...8 °C in bag with desiccant	6 weeks
Control	reconstituted	-18 °C or below	6 weeks
Calibrators	reconstituted	-18 °C or below	6 weeks
Substrate Buffer	opened	2...8 °C	6 weeks
Wash Buffer	diluted	2...8 °C	6 weeks
Sample Diluent	opened	2...8 °C	6 weeks
Releasing Reagent	opened	2...8 °C	6 weeks
Stop Solution	opened	2...8 °C	6 weeks

Do not use the reagents after the expiry date.

Note:

The specimens' storage and stability information stated above are general recommendations for use in a variety of settings of laboratories. Each laboratory should follow the guidelines or requirements of local, state, and/or federal regulations or accrediting organizations to establish its own specimens handling and storage stability. For guidance on appropriate practices, please refer to the CLSI GP44-A4, Procedures for the Handling and Processing of Blood Specimens for Common Laboratory Tests; Approved Guideline - Fourth Edition

2. REAGENTS AND MATERIALS REQUIRED BUT NOT PROVIDED

- 2.1. Deionised (DI) water.
- 2.2. Adjustable micropipettes.
- 2.3. Clean glass or plastic containers for dilution of Wash Buffer and specimen.
- 2.4. Suitable device for microplate washing (e.g. multistepper or ELISA washer).
- 2.5. Incubator for 37 °C.

2.6. Microplate shaker, shaking frequency 850–950 rpm, amplitude 4 mm.

2.7. Microplate reader with filter for 405 nm.

3. PREPARATION OF THE REAGENTS

Before starting the test procedure all kit components must be equilibrated to room temperature.

Calculate the number of wells required.

3.1. Microplate

The aluminium bag has to be tightly resealed together with the desiccant after each removal of wells. Storage and stability of the wells are indicated in table 1.

3.2. Wash Buffer

Mix one volume of Wash Buffer (25x) with 24 volumes of water for injection (e.g. 10 ml Wash Buffer (25x) with 240 ml water). Seven ml of diluted wash buffer is needed for 8 wells.

3.3. Calibrators

Reconstitute the lyophilised calibrators each with 0.5 ml of water for injection. Reconstitution time 15 min.

3.4. Controls

Reconstitute the lyophilised controls with 0.5 ml of water for injection. Reconstitution time 15 min.

3.5. Substrate Solution

1 Substrate Tablet is dissolved in 5 ml Substrate Buffer.

The Substrate Solution must not be stored.

Do not mix reagents from different lots or manufacturers.

Valid and reproducible results are only obtained if the test procedure is precisely followed and test kit-specific reagents are used.

4. SPECIMEN

4.1. The assay is suitable for serum and EDTA-plasma samples.

NB. The same specimen type must be used throughout a follow-up study.

4.2. Pretreatment of sera, e.g. inactivation, must not be performed. The specimen should not be contaminated with microorganisms.

- 4.3. Specimen can be stored up to 8 hours at room temperature and up to 3 days at 2-8 °C. Storage at –20 °C is possible for 2 months. For longterm storage a temperature of –80 °C is necessary.
- 4.4. Samples are used undiluted. Specimen above the measuring range can be diluted up to 1:5.

5.A. TEST PROCEDURE

- 5.1. Cut the aluminium bag above the zip fastener and take out the required number of microplate wells (see 3.1.).

Microplate wells are ready to use and do not have to be pre-washed.

NB! Microplate 12x8 wells.

- 5.2. Add 100 µl each of Calibrators, Control and samples to the wells of the plate in duplicate.
- 5.3. Add 50 µl Releasing Reagent to each well.
- 5.4. Seal the microplate with incubation cover foil and incubate for 60 min (\pm 5 min) at room temperature with constant shaking at 850–950 rpm.
- 5.5. After incubation wash the microplate wells four times with 300 µl wash buffer per well. Pay attention that all wells are filled. After washing tap microplate wells on filter paper.

Do not allow the wells to dry out! Proceed immediately!

- 5.6. Add 100 µl Substrate Solution to each well.
- 5.7. Seal the microplate with incubation cover foil and incubate for 60 min (\pm 5 min) at 37 °C (\pm 1 °C).
- 5.8. Stop the reaction by adding 25 µl of Stop Solution to each well.

Ensure for a good mixing by shaking gently.

Clean microplate wells from underneath before the photometric reading and take care that there are no air bubbles in the wells.

The reading should be done within 15 min after adding the Stop Solution!

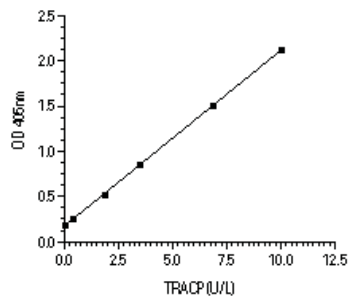
5.B. TABLE FOR THE TEST PROCEDURE

	Calibrators	Control	Sample
Calibrators	100 µl	-	-
Controls	-	100 µl	-
Sample	-	-	100 µl
Releasing Reagent	50 µl	50 µl	50 µl
Incubate for 60 min (± 5 min) at room temperature with constant shaking at 850–950 rpm, wash 4 x with 300 µl wash buffer.			
Substrate Solution	100 µl	100 µl	100 µl
Incubate for 60 min (± 5 min) at 37 °C (± 1 °C).			
Stop Solution	25 µl	25 µl	25 µl
Photometric reading at 405 nm			

6.A. CALCULATION OF RESULTS (VALIDITY)

- * Read OD values at 405 nm.
- * The average OD values of the calibrators are plotted against the activity values. The calibration line is calculated by linear regression.
- * The measuring range spans from 0.5 to 10 U/L. Samples below the measuring range have to be interpreted as < 0.5 U/L. Samples with activities above the measuring range have to be interpreted as > 10 U/L. These values must not be extrapolated but the samples should be retested diluted (up to 1:5).
- * The TRACP 5b activities of the controls and the samples can be read from the calibration line. If diluted samples have been used the dilution factor has to be considered.

Example for calibration line:



- * Lot-specific data
Refer to Quality Control Report.
- * Validity criteria
 - The average OD of the 0 U/L calibrator has to be < 0.400.
 - The activity of the control has to be within the nominal range. Refer to Quality Control Report for assigned range.
 - The correlation coefficient (r^2) of the calibration line has to be ≥ 0.99 .

Repeat the run if the results do not meet the specification!

6.B. INTERPRETATION OF RESULTS/LIMITATIONS OF THE METHOD

- * Increased TRACP 5b activity (see 7.E.) indicates an increased bone resorption.
- * Results within the reference range do not exclude disorders in bone metabolism completely. Therefore all results should always be interpreted in connection with clinical data and additional diagnostic parameters.
- * High concentrations of hemoglobin do not influence the test results.
- * High lipid concentrations may reduce the OD values and distort the TRACP activity.

7. PERFORMANCE CHARACTERISTICS

We determined the following performance characteristics during the evaluation of the assay.

7.A. PRECISION

Sample	Intra-assay variation (n = 21)			Sample	Inter-assay variation (n = 11)		
	mean U/L	SD	CV (%)		mean U/L	SD	CV (%)
Control	3.0	0.18	6.0	Control	3.3	0.19	5.8
N° 1	2.6	0.25	9.6	N° 1	2.5	0.23	9.2
N° 2	3.1	0.43	13.9	N° 2	4.2	0.35	8.3
N° 3	7.1	0.47	6.6	N° 3	7.0	0.62	8.9
				N° 4	7.2	0.38	5.4
				N° 5	2.6	0.23	8.8
				N° 6	16.1	1.16	7.2

7.B. RECOVERY

By adding 3 defined TRACP 5b activities each to 3 different sera a mean recovery of 100.9 % (SD = 11.3 %) was determined.

7.C. DILUTION LINEARITY

A linearity was determined by using sera of different activity (n=5). Samples with high TRACP 5b activity can be diluted up to 1:5 with Sample Diluent.

7.D. LIMIT OF QUANTITATION

The limit of quantitation is < 0.5 U/L.

7.E. EXPECTED VALUES

Expected values of TRACP 5b were determined from the serum of 239 healthy blood donors as follows:

Group	n	Mean Age (range)	TRACP 5b (U/L) Mean \pm SD
Healthy premenopausal women	144	39.5 (22 – 54)	2.59 \pm 0.78
Healthy young men	32	36.0 (22 – 54)	3.06 \pm 0.88
Healthy postmenopausal women	46	60.3 (41 – 81)	3.19 \pm 0.85
Healthy old men	17	68.5 (55 – 79)	3.31 \pm 0.72

Upper limits of normal were calculated as the mean + 2 SD of premenopausal women (for women) and young men (for men):

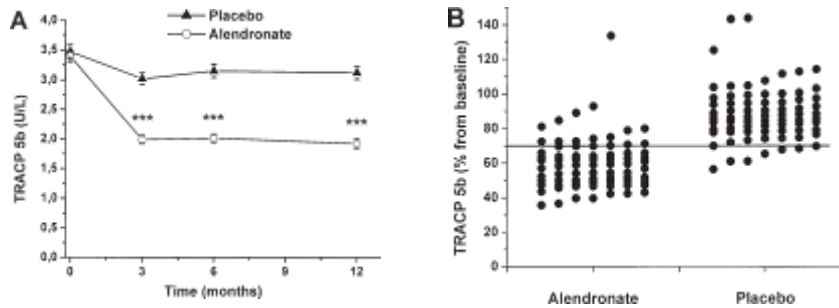
Group	Upper normal limit (mean + 2SD)
Women	4.15 U/L
Men	4.82 U/L

8. CLINICAL IMPORTANCE

TRACP 5B ACTIVITY IN POSTMENOPAUSAL WOMEN UNDER ALENDRONATE THERAPY

(see ref. 14)

TRACP 5b values were determined from the serum of postmenopausal women receiving 5 mg alendronate daily for 12 months in a placebo-controlled study. All subjects in the placebo (n = 73) and alendronate (n = 75) groups received a daily supplement of 630 mg calcium carbonate and 200 IU vitamin D.



A) Serum TRACP 5b activity (U/L) before the start of treatment (0) and at 3, 6 and 12 months; B) The change of serum TRACP 5b at 3 months. Each spot shows the value of one individual at 3 months compared with the value obtained for the same individual at baseline (before the start of treatment). The line in B) shows the Least Significant Change (LSC = 29.5%). A decrease of more than LSC was observed for 82.7% of the individuals in the alendronate group, and 11.0% of the individuals in the placebo-group.

GENERAL HANDLING ADVICES

- * To avoid cross contamination do not exchange the vials and their screw caps.
- * The reagents have to be sealed immediately after use to avoid evaporation and microbial contamination.
- * After use, the reagents have to be stored as indicated to guarantee the shelf life.
- * After use, all components of the testkit should be stored in the original package, in order to avoid mixing up the reagents of other test systems or lots (see also 3.).

HEALTH AND SAFETY INFORMATION

- * The local occupational safety and health regulations have to be regarded.
- * Reagents of animal origin (see kit contents) should be handled as potentially infectious and used with all necessary precautions.

Classification under CLP:

Acute Tox. 4:

Aquatic Chronic 3:

EUH032

Hazard statements:

EUH032: Contact with acids liberates very toxic gas.

H302: Harmful if swallowed.

H315: Causes skin irritation.

H319: Causes serious eye irritation.

H412: Harmful to aquatic life with long lasting effects.

Precautionary statements:

P264: Wash hands thoroughly after handling.

P280: Wear protective gloves/protective clothing/eye protection/face protection.

P270: Do not eat, drink or smoke when using this product.

P301+310: IF SWALLOWED: Immediately call a POISON CENTER or doctor.

P305+351+338: IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

P330: Rinse mouth

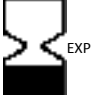




Wash Buffer, Sample Diluent, Releasing Reagent and Substrate Buffer contain Germall®II (diazolidinyl urea): May produce an allergic reaction.

DISPOSAL CONSIDERATIONS

Residues of chemicals and preparations are generally considered as hazardous waste. The disposal of this kind of waste is regulated through national and regional laws and regulations. Contact your local authorities or waste management companies which will give advice on how to dispose hazardous waste.

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